



anses

***VIBRIO AND AEROMONAS IN FRESHWATER:
A FOCUS ON V. CHOLERAЕ AND A. SALMONICIDA
TWO « ZONOTIC » PATHOGENS?***

SANDRINE BARON

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NIORT LABORATORY***

CONNAÎTRE, ÉVALUER, PROTÉGER

Why *Vibrio cholerae* & *Aeromonas salmonicida*?

autochthonous of aquatic environment

Gram negative, oxydase +



Available online at www.sciencedirect.com
ScienceDirect

Aeromonas: the multifaceted middleman in the One Health world
Brigitte Lamy^{1,2,3}, Sandrine Baron⁴ and Olivier Barraud⁵

RESEARCH ARTICLE
15 JULY 2015 | Volume 24, Number 142

Agreement between the categorization of isolates of *Aeromonas salmonicida* and *Yersinia ruckeri* by disc diffusion and MIC tests performed at 22°C

Sandrine Baron¹ | Emeline Larvois² | Eric Jouy³ | Isabelle Koron⁴
Sophie Le Douarin¹ | Claire Charrier¹ | Pierre-Martin Gobard⁴ |
Alain Le Breton¹ | Denoit Thuiller² | Peter Smith⁵ | Matthieu Jamin⁵

ELSEVIER
WILEY

Antimicrobial Susceptibility among Urban Wastewater and Wild Shellfish Isolates of Non-O1/Non-O139 *Vibrio cholerae* from La Rance Estuary (Brittany, France)
Sandrine Baron^{1,2}, Stéphanie Lecerf^{1,2}, Stéphanie Chevalier^{1,2}, Eric Jouy³, Isabelle Koron⁴, Sophie A. Gervais^{1,2} and Jean Lecaillon^{1,2}

***Vibrio cholerae* in the Environment: A Simple Method for Reliable Identification of the Species**
S. Baron, S. Chevalier, and J. Lecaillon
Laboratoire d'Étude et de Recherche en Environnement et Santé,
École Nationale de la Santé Publique, Rennes, France



La pisciculture – STEB (aquaculteurs-de-bretagne.fr)



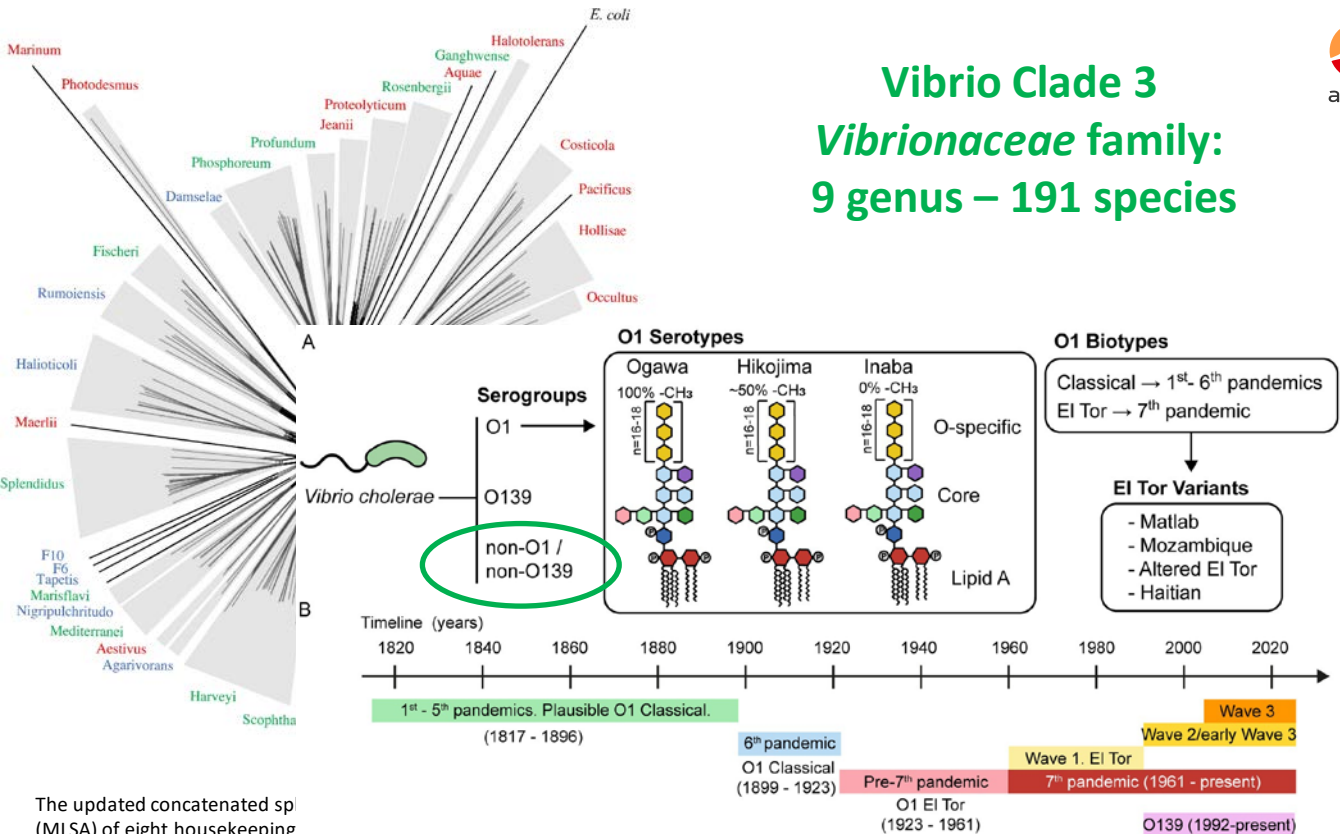
Four decorative orange leaf-like shapes are scattered across the slide, partially overlapping the text. They are positioned at approximately (520, 240), (710, 240), (500, 500), and (650, 600) in normalized coordinates.

Complex and constantly evolving taxonomy

Vibrio Clade 3

Vibrionaceae family:

9 genus – 191 species



The updated concatenated sp (MLSA) of eight housekeeping species. The *ftsZ*, *gapA*, *gyrB*, *mreB*, *pyrH*, *recA*, *rpoA*, and *topA* gene sequences were concatenated and the tree was reconstructed using the SplitsTree4 ver. 4.14.8. Clades indicated by red, green, and blue represent the “new,” “emended,” and “unchanged” clades, respectively

36 species of *Aeromonas*

5 subspecies of *Aeromonas salmonicida* (*A. sal*):

A. sal. subsp. salmonicida – psychrophilic*

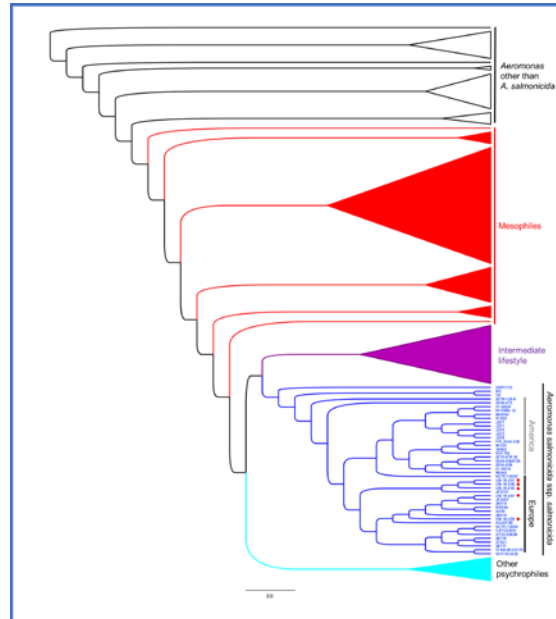
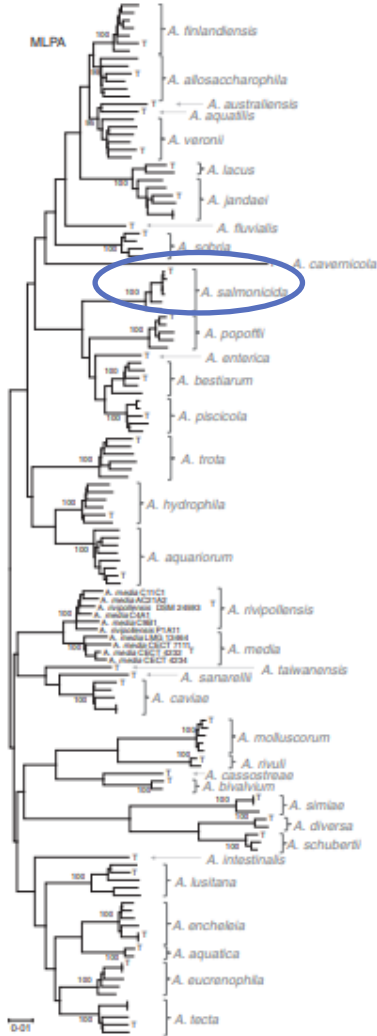
A. sal. subsp. achromogenes - psychrophilic **

A. sal. subsp. masoucida - psychrophilic**

A. sal. subsp. pectinolytica - mesophilic

A. sal. subsp. smithia - psychrophilic**

+ myriad of unclassified strains



**

Complex and constantly evolving taxonomy...

↪ Difficulties of identification

↪ Ecological meaning of the « new described » species?

↪ Impact in Human & Animal Health?



Ecotype (Fred Cohan 2002) ?

Eukaryote systematists have developed a universal concept of species: A species is a group of organisms whose divergence is capped by a force of cohesion; divergence between different species is irreversible; and different species are ecologically distinct. In the case of bacteria, these universal properties are held not by the named species of systematics but by ecotypes. **These are populations of organisms occupying the same ecological niche, whose divergence is purged recurrently by natural selection.**

Can *V. cholerae* and *Aeromonas* spp
be considered as candidates to be potential indicators of
antimicrobial resistance dissemination
in the aquatic environment?



Criteria to be fulfilled:

- An easy and frequent detection in aquatic environment
- Accurate and rapid identification tools
- A capacity to acquire and exchange genes
- Cut-off values to categorize the isolates between wild type and non-wild type.

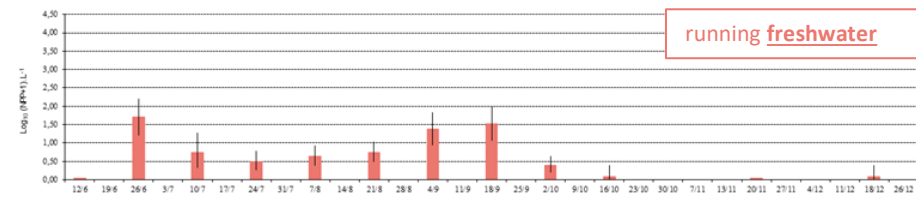
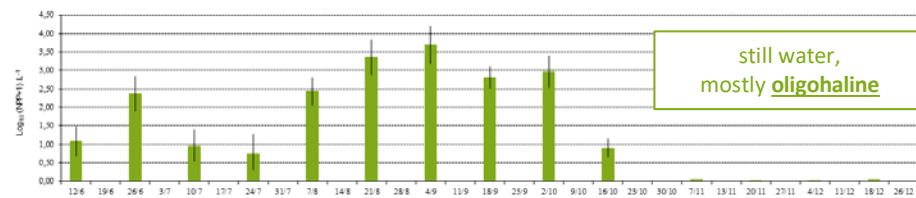
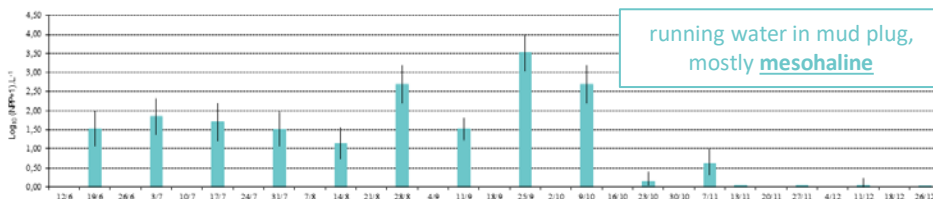
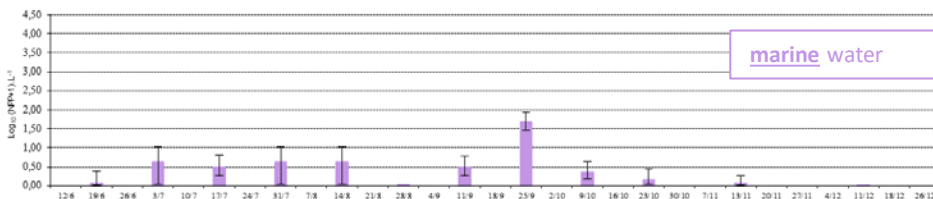
Vibrio cholerae – *Aeromonas* spp.



An easy and frequent detection in aquatic environment: *V. cholerae*

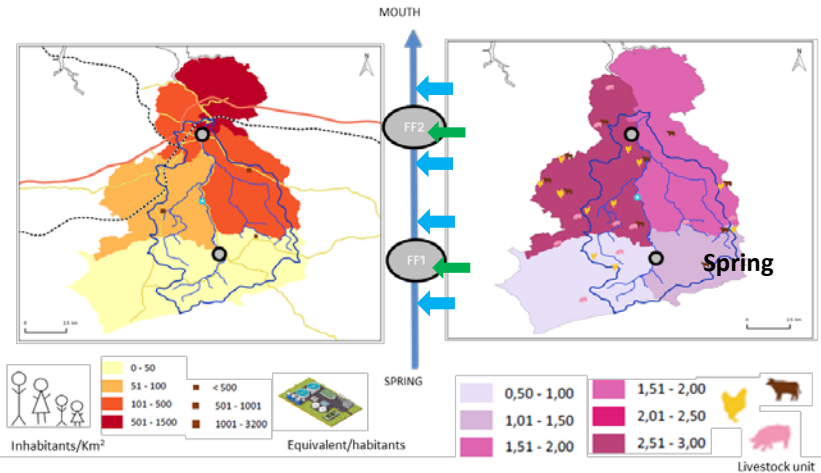


V. c abundances between June – December en $\log_{10}(\text{MPN}+1) \cdot \text{L}^{-1}$



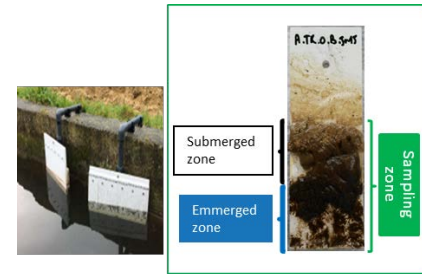
An easy and frequent detection in aquatic environment:

Aeromonas spp.



South-North gradient of anthropization:

humane density & agricultural activities three wastewater treatment plans



Device for biofilm sampling in ponds

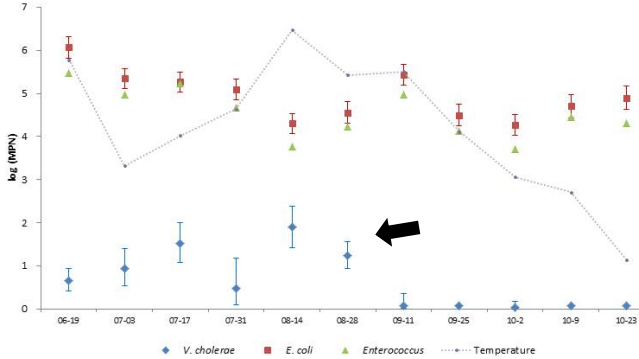
Aeromonas abundances during 17 months (CFU/mL)

	Nb samples	Prevalence ¹	Abundances ²	
Up. FF1	36	100%	10 ⁰	10 ³
Biofilm FF1	36	100%	>10 ³	>10 ⁶
Down. FF1	36	100%	10 ¹	10 ⁴
Up. FF2	36	100%	10 ¹	10 ³
Biofilm FF2	36	100%	>10 ³	>10 ⁶
Down. FF2	36	100%	10 ¹	10 ³

¹positive samples, ²Min – Max (CFU/mL), WW: wastewater, FF: Fishfarm

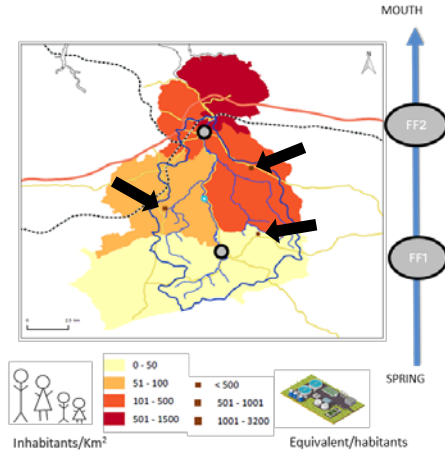
Detection in raw and treated wastewater ?

Vibrio cholerae (MPN/L): June-December



Aeromonas spp. (CFU/mL): January - June

	Nb samples	Prevalence	Abundances Min – Max
Raw 1	3	100%	>10 ⁵ >10 ⁵
Treated 1	3	100%	>10 ³ >10 ⁵
Raw 2	5	100%	>10 ² >10 ⁵
Treated 2	5	100%	>10 ³ >10 ⁵
Raw 3	5	100%	>10 ⁴ >10 ⁵
Treated 3	5	100%	>10 ² >10 ³



An easy and frequent detection in aquatic environment

Vibrio cholerae: easy – period of no detection

easy – detection all year long

Aeromonas

Accurate and rapid identification tools

A capacity to acquire and exchange genes

Cut-off values to categorize the isolates between wild type and non-wild.

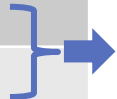


Accurate and rapid identification tools

	PCR	Maldi-ToF
Genus <i>Vibrio</i>	no	yes
Species of <i>Vibrio</i>	-	(no)
<i>Vibrio cholerae</i>	yes	yes
Genus <i>Aeromonas</i>	yes	yes
Species of <i>Aeromonas</i>	-	no
<i>Aeromonas salmonicida</i>	yes	no



Collaboration with BFR

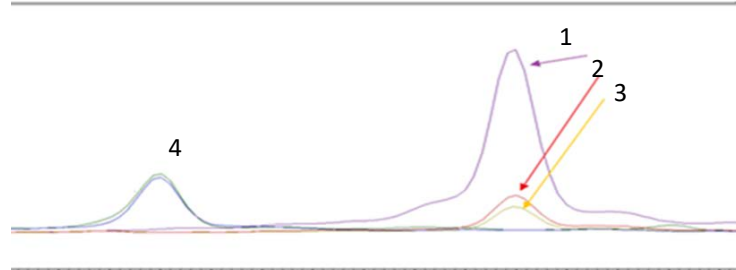


Almost ready ...

↪ Improvement of database (Maldi-ToF)

Species	n	Bruker database		anses database	
		DD	DA	DD	DA
<i>Aeromonas bestiarum</i>	68	91%	96%	99%	99%
<i>Aeromonas popoffii</i>	20	5%	0%	70%	80%
<i>Aeromonas rivuli</i>	1	0%	0%	100%	100%
<i>A. salmonicida</i>	78	87%	78%	83%	96%

↪ Detection of biomarkers (Maldi-ToF)



1 *A. sobria*, 2 *A. bestiarum*,
3 *A. enterica*, 4 *A. salmonicida* & *A. enterica*

An easy and frequent detection in aquatic environment

Vibrio cholerae: ease - not detection period

Aeromonas spp: ease – detection all the year

Accurate and rapid identification tools

Vibrio cholerae: PCR & Maldi-ToF

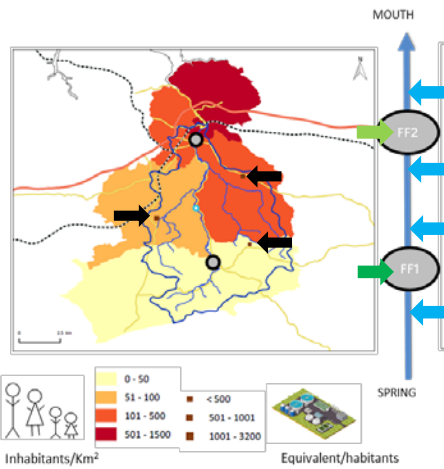
Aeromonas spp: PCR & Maldi-ToF

A capacity to acquire and exchange genes

Cut-off values to categorize the isolates between wild type and non-wild.

Aeromonas spp. In water & biofilm of fishfarm: Integron

601 isolates – 98 harbored integron (16,3%)



	Nb samples	Positive samples	Nb isolates	% integron
S1	18	100%	85	1.2% (1)
S6	18	100%	87	27.6% (24)
Raw WW1	3	100%	30	3.3% (1)
Treated WW1	3	100%	30	10.0% (3)
Raw WW2	5	100%	49	2.0% (1)
Treated WW2	5	100%	49	6.1% (3)
Raw WW3	5	100%	48	25.0% (12*)
Treated WW3	5	100%	48	6.25% (3)
Biofilm FF1	18	61.1%	87	24.4% (21)
Biofilm FF2	18	66.7%	88	32.9% (29)

- Increase of integron in water from spring to mouth (X23)
- Average in raw wastewater: 11.6%, in treated wastewater: 12.6% (2% -25%)
- No difference in Biofilm

Aeromonas & fish: Integron

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Aquaculture

journal homepage: www.elsevier.com/locate/aquaculture



Integrations from *Aeromonas* isolates collected from fish: A global indicator of antimicrobial resistance and anthropic pollution

Olivier Barraud^{a,1}, Lucie Laval^{a,1}, Laëtitia Le Devendec^b, Emeline Larvor^b, Claire Chauvin^c, Eric Jouy^b, Sophie Le Bouquin^c, Yann Vanrobaeys^b, Benoit Thuillier^d, Brigitte Lamy^{e,f}, Sandrine Baron^{b,4}

Integrations detected from *Aeromonas* collections.

Fish category (number of isolates)	<i>Aeromonas salmonicida</i> (n = 39)		<i>Aeromonas</i> spp. non <i>salmonicida</i> (n = 340)		All <i>Aeromonas</i> (n = 379)	
	Integron positive	Integron negative	Integron positive	Integron negative	Integron positive	Integron negative
Wild Fish – WF (n = 78)	0	0	0	78	0	78
Farmed fish – FF (n = 231)	0	0	35	196	35	196
Diseased fish – DF (n = 70)	25*	14	9	22	34*	36
Total (n = 379)	25*	14	44	296	69*	310

*Includes the only class 2 integron.

Highest frequencies of integron in *A. salmonicida* isolates

A. salmonicida isolates in diseased fish and farmed fish

A capacity to acquire and exchange genes: mobile genetic element

Aeromonas & fish: Integron



Minimum inhibitory concentration (MIC) probability density, box plot and scatter plot by fish category

WF: wild fish; FF: farmed fish; DF: diseased fish

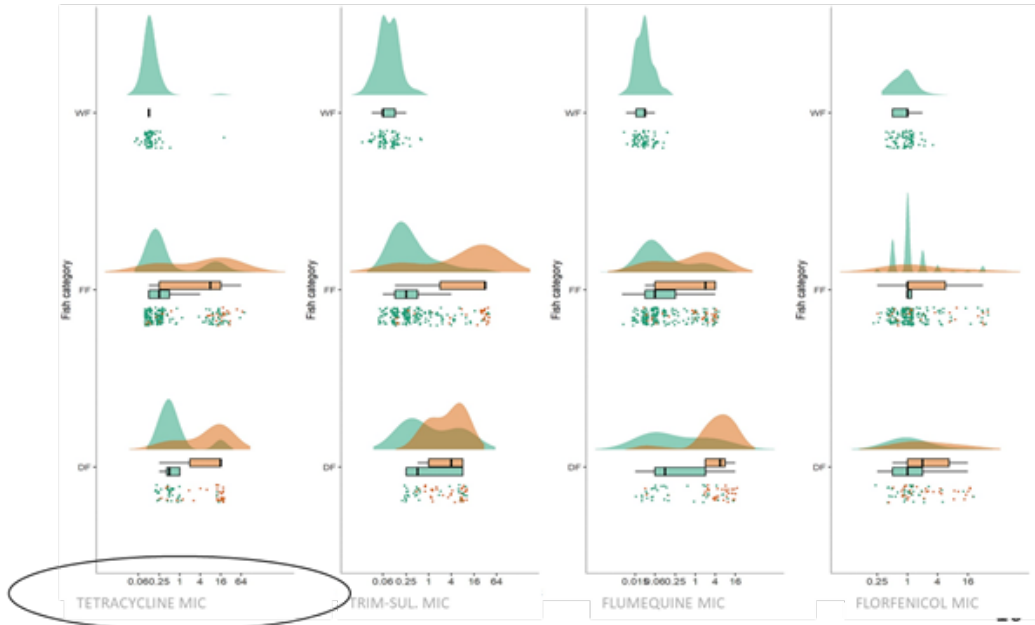
- according to integrons **presence** (orange) or **absence** (green)

FISH CATEGORY

Wild
n = 78

Farmed
n = 231

Diseased
n = 70

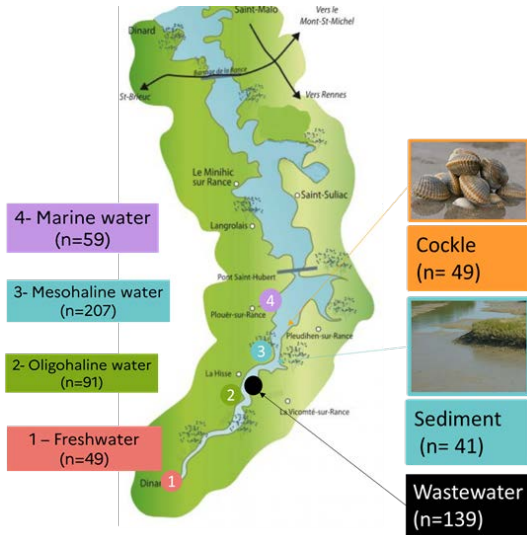


Integron
negative

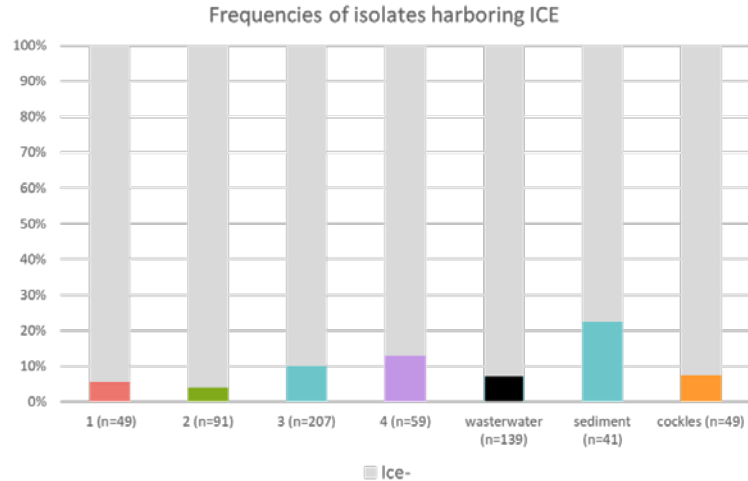
Integron
positive

↪ Lowest MIC for isolates **without integron**
↪ Difference between « fish » population

Vibrio cholerae: Integrative Conjugative Element



The 635 isolates were subjected to a molecular survey on the presence of ICE



ICE of the SXT/R391 family was detected in 10.9% of the analyzed strains (n=64)

V. cholerae isolation was conducted from water, sediment and cockles samples collected along a salinity gradient under a temperate climate (France) between June 2000 and September 2001

isolates carrying ICE were further subjected to paired-end, short read whole-genome sequencing (WGS) for in depth characterization (BfR) ...

An easy and frequent detection in aquatic environment

Vibrio cholerae: ease - not detection period

Aeromonas spp: ease – detection all the year

Accurate and rapid identification tools

Vibrio cholerae: PCR & Maldi

Aeromonas spp: PCR & Maldi

A capacity to acquire and exchange genes

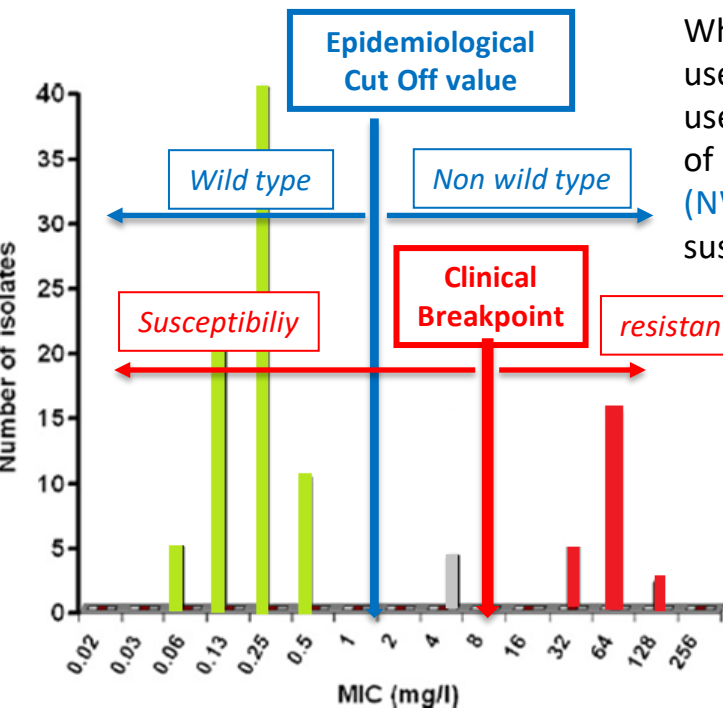
Vibrio cholerae: ICE

Aeromonas spp: INTEGRON

Cut-off values to categorize the isolates between wild type and non-wild.

Epidemiological Cut-off values to categorize the isolates between wild type and non-wild.

Clarification of the appropriate terminology by Silley (2012)



When **Epidemiological Cut Off value** are used the term **Wild-type (WT)** should be used to refer to fully **susceptible** members of a species and the term **non-wild Type (NWT)** to isolates that manifest a reduced susceptibility

Resistance should only be used to refer to **clinical resistance**.

Terms **resistant** and **sensitive** should only be used when susceptibility data has been interpreted by application of **Clinical Breakpoint**

Vibrio cholerae non-O1/non-O139

Epidemiological cut-off values for non-O1/non-O139 *Vibrio cholerae* disc diffusion data generated by standardised methods

Peter Smith*, Laëtitia Le Devendec, Eric Jouy, Emeline Larvor, Jean Lesne, Alexander K. T. Kirschner, Carmen Rehm, Melanie Leopold, Sonja Pleininger, Florian Heger, [Claudia Jäckel](#), [Cornelia Göllner](#), [Jonas Nekat](#), [Jens Andre Hammerl](#), Sandrine Baron

Agent	CO _{WT} (mm)	S (mm)		Difference (CO _{WT} - CB)
		M45 ^a	M100 ^b	
Aminoglycosides				
Amikacin	≥18	≥17 ^c		1
Gentamicin	≥16	≥15 ^c		1
Streptomycin	≥13		≥15	-2
Aminopenicillins				
Ampicillin	≥19	≥17		2
Amoxicillin/ clavulanate	≥18	≥18 ^c		0
Carbapenems				
Meropenem	≥25	≥23 ^c		2
Imipenem	≥23	≥23 ^c		0
Cephems				
Cefotaxime	≥31	≥26 ^c		5
Ceftazidime	≥24	≥21 ^c		3
Cefepime	≥27	≥25 ^c		2
Folate pathway inhibitors				
Trimethoprim-sulfamethoxazole	≥24	≥16		8
Trimethoprim	≥23		≥16	7
Macrolides				
Erythromycin	≥16	na	na	
Phenicol				
Chloramphenicol	≥24	≥18		5
Florfenicol	≥27	na	na	
Quinolones				
Norfloxacin	≥28		≥17	11
Ciprofloxacin	≥31	≥21		10
Nalidixic acid	≥29		≥19	10
Tetracyclines				
Tetracycline	≥23		≥15	8

Publication in press

Aeromonas spp.

frontiers
in Microbiology

ORIGINAL RESEARCH
published: 20 March 2017
doi: 10.3389/fmicb.2017.00505



Aeromonas Diversity and Antimicrobial Susceptibility in Freshwater—An Attempt to Set Generic Epidemiological Cut-Off Values

Sandrine Baron^{1,2*}, Sophie A. Granier¹, Emeline Larvor^{1,2}, Eric Jouy^{1,2}, Maëlan Cizeux^{1,2}, Amandine Wilhelm¹, Benoit Gassilloud¹, Sophie Le Bouquin^{1,2}, Isabelle Kempf^{1,2} and Claire Chauvin^{1,2}

Provisional Ecoff for 12 antibiotics.

Aeromonas salmonicida (in press).

Agent ^a	IZ data (mm)			
	AMP	FLO	OXO	OXY
VET 04 ^b	na	≥27	≥30	≥28
This work ^c	≥29	≥30	≥32	≥25

Agent ^a	MIC data (µg/ml)		
	FLO	OXO	OXY
VET 04 ^b	≤4	≤0.125	≤1
This work ^c	≤4	≤0.125	≤1

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Vibrio cholerae: ease - not detection period

Aeromonas spp: ease – detection all the year

Accurate and rapid identification tools

Vibrio cholerae: PCR & Maldi

Aeromonas spp: PCR & Maldi

A capacity to acquire and exchange genes

Vibrio cholerae: ICE

Aeromonas spp: INTEGRON

Cut-off values to categorize the isolates between wild type and non-wild type

Vibrio cholerae: 19 antibiotics

Aeromonas spp & *A. salmonicida*: 12 & 4 antibiotics

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Vibrio cholerae: 19 antibiotics

Aeromonas spp & *A. salmonicida*: 12 & 4 antibiotics

Vibrio cholerae << *Aeromonas spp*

BUT - Intrinsic resistance in *Aeromonas*: C3G & carbapenem

- Diversity of species

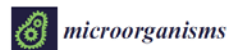
AND *Vibrio cholerae* susceptible to C3G and carbapenem



Can they be considered as zoonotic pathogen?

V. cholerae - *Aeromonas salmonicida*: zoonotic pathogen?

With climate change:
increase of density in water
increase of humane cases



Article
**Phenotypic and Genotypic Properties of
Vibrio cholerae non-O1, non-O139 Isolates
Recovered from Domestic Ducks in Germany**

Nicola Hirsch¹, Eva Kappe¹, Armin Gangl¹, Keike Schwartz², Anne Mayer-Scholl²,
Jens Andre Hammerl² and Eckhard Strauch²

Infection due to:

V. c non-O1/non-O139



Also in goat, lambs, horse,
american buffalo ...

A. salmonicida



Mesophilic one* ...



Detection in « healthy »...

V. c non-O1/non-O139-VIM-1 and VIM-4
yellow-legged gull in southern France

Aquatic animal, terrestrial animal

A. salmonicida

Aquatic animal, pig**



Contamination between animal species:



Great egret*



catfish

tilapia

Great cormorant



Collaboration BfR-Anses

Vibrio spp – *Aeromonas spp*: Phage & Aquaculture



In BfR
Nov 2022



In Anses
May 2023



Short-Term Mission

In 2024 STM in BfR and STM in Anses . . .



Short-Term Mission



anses

DANKE ...

SANDRINE BARON

***MYCOPLASMOLOGY-BACTERIOLOGY AND ANTIMICROBIAL RESISTANCE UNIT PLOUFRAGAN-PLOUZANÉ-
NIORT LABORATORY***

CONNAÎTRE, ÉVALUER, PROTÉGER